Imaging pulsatile retinal blood flow in human eye

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Abstract. A functional Fourier domain optical coherence
tomography instrument offering spectral Doppler imaging
of in vivo pulsatile human retinal blood flow was con-
structed. An improved phase-resolved algorithm was de-
vloped to correct bulk motion artifacts. Spectral Doppler
imaging provides complementary temporal flow informa-
tion to the spatially distributed flow information of the
color Doppler image by providing direct visualization of
the Doppler spectrum of the flow whose pattern can be
further quantified with various velocity envelope curves
and their corresponding flow indices. The coefficient of
repeatability on resistance index measurement was as-
sessed by analyzing 14 measurements on two vessels
within two normal subjects.© 2008 Society of Photo-Optical In-
strumentation Engineers. [DOI: 10.1117/1.2967986]

Keywords: doppler; tomography; interferometry.

JBO LETTERS
Journal of Biomedical Optics July/August 2008 /Vol. 13/No. 4

Because of the unknown Doppler angle between the blood
flow and incident light beam and the absence of a quantifica-
tion method that can generate interpretable results for cli-
nicians, DOCT is not widely used in eye clinics. Conventional
DOCT only generates a snapshot of pulsatile ocular blood
flow that is projected along the light beam direction in a
cardiac cycle. Most recent developments in quantifying blood
flow information of the human eye utilizes 3-D vascular ori-
entation information to estimate Doppler angle and the abso-
late flow velocity. On the other hand, it is noteworthy to
quantify the pulsatile flow pattern as an alternative method to
investigate retinal flow dynamics. A simple projected ocular
blood flow velocity (integration over the whole blood vessel)
plot through a cardiac cycle was chosen by White to dem-
strate the pulsatile flow property, although it is possible to
acquire much more hemodynamics information from the same
raw data. The M-mode scanning method has been used to
acquire temporal flow information in time-domain DOCT
systems. The short-time fast Fourier transformation
method from Doppler ultrasound was used to generate Dop-
pler spectrum wave forms but without further quantification,
which provides the most valuable information for clinicians.
The purpose of this paper is to implement the full concept of
spectral Doppler imaging, developed by scientists and clini-
cians in ultrasound medicine, in a Fourier-domain DOCT sys-
tem and provide an alternative quantification method for an
ocular blood flow pattern that may be further investigated for
vascular related eye diseases.

“Spectral Doppler” is a terminology from Doppler ultra-
sound and should not be related to spectral OCT that uses
spectral information as a contrast mechanism of OCT. Spectr-
Al Doppler imaging of pulsatile retinal blood flow includes
Doppler spectrum visualization, using spectral Doppler wave
forms, and a method for quantifying the temporal properties
of flow, using various velocity envelopes and their corre-
sponding Doppler-angle-independent indices. Continuous
color Doppler data are acquired when the light beam performs
repeated dense scans over the region of interest. Spectral Dop-
pler analysis on the data shows how the velocity components
and longitudinally projected flow-volume-rate change over
time for scatters within the imaging volume with spectral
Doppler wave forms. Various velocity envelope curves can be
derived from spectral Doppler wave forms and used to extract
the corresponding pulsatility index (PI), resistance index (RI)
and several other indices that can provide interpretable
Doppler-angle-independent information needed to quantify
the pulsatile nature of ocular blood flow.

A Fourier domain functional OCT system was developed
for retinal blood flow imaging. Briefly, low coherence light
with a center wavelength of 890 nm and FWHM bandwidth
of 150 nm was protected from optical feedback using a
broadband optical isolator before entering a 2 × 2 broadband
fiber coupler-based interferometer. The light from the refer-
cence arm was focused onto a reference mirror with an optical
attenuator inserted into the optical path. The sample arm was
modified from the patient module of a Zeiss Stratus OCT
instrument. The detection arm was connected to a high-
performance spectrometer that allows the system bench-top
sensitivity of 100 dB with 650 μW light out of the sample
arm fiber and 50 μs CCD integration. A 9 dB of SNR roll-off
from zero imaging depth to 2 mm imaging depth was ob-
served. The system has an axial resolution of 3.5 μm. In this
study, the system speed was set at 16.7 K A-lines/s with its
CCD A-line integration time set at 50 μs and the line period
set at 60 μs. The maximum longitudinal velocity in retinal
tissue that corresponded to a phase difference of π was deter-
m ined to be 2.69 mm/s according to

\[ V_{\text{max}} = \pi / (4T) \]

where \( T \) is the line period of the CCD camera. The measured phase
noise from a mirror was 2.66 mrad. The velocity measurement
error was determined to be <5% by imaging steady-

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1083-3668/2008/13(4)/040505/1/$25.00 © 2008 SPIE
state scattering flow (polystyrene bead solution with mean diameter of 0.3 μm and volume concentration of 0.26%) pumped at different velocities.

An improved phase-resolved algorithm that uses a Doppler variance image to select tissue pixels for histogram analysis of bulk phase was developed in this instrument to compensate for the axial eye movement. Most bulk motion presented in Fig. 1(a) has been corrected by the conventional histogram method in (c). An improved phase-resolved algorithm with more reduced artifacts than normal histogram method in (c). Spectral Doppler scan trace shown on 5×5 mm OCT fundus image (140 slices×512 A-lines), (f) the intensity and color Doppler image for one snapshot (700×256 pixels), (g) spectral Doppler wave forms that show the change of (axial) velocity and flow-volume-rate within a time span of 7.9 s (the right grayscale bar is used to represent the volume-rate contribution for a given velocity bin) and (h) the maximum velocity envelope curve of eight cardiac cycles from which the quantitative Doppler-angle-independent flow indices were calculated.

Table 1 Intrasession coefficient of variation of Doppler flow indices for one measurement.

<table>
<thead>
<tr>
<th>Cycle</th>
<th>PI</th>
<th>RI</th>
<th>SD</th>
<th>DA</th>
</tr>
</thead>
<tbody>
<tr>
<td>1</td>
<td>1.0753</td>
<td>0.6255</td>
<td>2.6709</td>
<td>0.64352</td>
</tr>
<tr>
<td>3</td>
<td>1.0551</td>
<td>0.621</td>
<td>2.6386</td>
<td>0.64393</td>
</tr>
<tr>
<td>4</td>
<td>1.1869</td>
<td>0.6805</td>
<td>3.1299</td>
<td>0.55727</td>
</tr>
<tr>
<td>5</td>
<td>1.1362</td>
<td>0.65217</td>
<td>2.875</td>
<td>0.606</td>
</tr>
<tr>
<td>6</td>
<td>1.4098</td>
<td>0.7561</td>
<td>4.1</td>
<td>0.45478</td>
</tr>
<tr>
<td>7</td>
<td>1.0667</td>
<td>0.61789</td>
<td>2.617</td>
<td>0.65965</td>
</tr>
<tr>
<td>8</td>
<td>1.0539</td>
<td>0.64</td>
<td>2.7778</td>
<td>0.59284</td>
</tr>
<tr>
<td>Mean</td>
<td>1.140557</td>
<td>0.656179</td>
<td>2.972743</td>
<td>0.593999</td>
</tr>
<tr>
<td>Std. Dev.</td>
<td>0.128572</td>
<td>0.049147</td>
<td>0.527933</td>
<td>0.070866</td>
</tr>
<tr>
<td>Coe. Var.</td>
<td>0.112727</td>
<td>0.0749</td>
<td>0.177591</td>
<td>0.119303</td>
</tr>
</tbody>
</table>

The assumption that the vessel has to be small compared to the tissues in the same A-line. For the scan protocol of spectral Doppler imaging, repeated color Doppler scans over the broken region of the red line shown in Fig. 1(e) for a short period of time were performed after a specific vessel was selected from the OCT fundus image generated from a 3-D scan immediately before spectral Doppler imaging. Spectral Doppler analysis was performed on the color Doppler images. In order to evaluate variation of measurements within one session, spectral Doppler imaging of 512 snapshots through eight cardiac cycles in 7.9 s was performed on one normal subject after the vessel was selected as shown in Fig. 1(e). Each snapshot used 256 A-lines on 132.5 μm tissue. The imaging speed of 65 snapshots per second is fast enough to capture the dynamic flow during a cardiac cycle. The structure image and velocity image for a typical snapshot are shown in Fig. 1(f). The longitudinal velocity sensitivity was 174 μm/s by fitting the velocity profile and calculating the standard deviation for the velocity image across the center of the vessel in Fig. 1(f).

Before spectral Doppler analysis, a threshold was applied to the Doppler phase image and then some morphological operations were performed to get a vessel mask. The vessel center could then be easily determined from the peak positions after projecting the vessel mask to horizontal and vertical directions. A rectangular window centered at the estimated vessel center position can be applied to the velocity image in Fig. 1(f).

Next, the spectral Doppler wave forms in Fig. 1(g) could be generated after spectral Doppler analysis was performed for every velocity image snapshot that corresponded to one vertical line of the spectral Doppler wave forms. The velocity range was digitized into 256 velocity bins (v0−v255). An iteration of all the pixels within the window generates a function \( n_i(v_i) \) that represents the number of pixels having velocity of \( v_i \). Because each pixel represents a vortex for the vessel lumen area, the product of the velocity of the velocity bin and the number of pixels that fall within the velocity bin produced gray scale amplitude, which is proportional to the longitudinal

Fig. 1 (a) Doppler image without bulk phase correction, (b) Doppler image with bulk phase correction by the conventional histogram method, (c) enlarged Doppler image of a selected area within the red window in (b), (d) doppler image generated by the improved phase-resolved algorithm with more reduced artifacts than normal histogram method in (c), (e) Spectral Doppler scan trace shown on 5×5 mm OCT fundus image (140 slices×512 A-lines), (f) the intensity and color Doppler image for one snapshot (700×256 pixels), (g) spectral Doppler wave forms that show the change of (axial) velocity and flow-volume-rate within a time span of 7.9 s (the right grayscale bar is used to represent the volume-rate contribution for a given velocity bin) and (h) the maximum velocity envelope curve of eight cardiac cycles from which the quantitative Doppler-angle-independent flow indices were calculated.
projection of the flow-volume-rate contributed by each velocity bin. The summation of the gray scale amplitude along the y-axis gives the total longitudinal projection of flow-volume-rate of a vessel at a given time point. The summation of the gray scale amplitude along the y- and x-axes for one cardiac cycle provides the longitudinal projection of total flow volume within that cardiac cycle. Note that spectral Doppler wave forms with other gray scale intensity definitions can be defined according to other physical meanings, such as particle numbers or particle energies. Different color channels may be introduced to represent multiple flow-related parameters in a color-coded plot of spectral Doppler wave forms.

In order to obtain more quantitative, interpretable results, different Doppler velocity envelopes, such as maximum velocity envelope, mean velocity envelope, and flow-volume-rate envelope, can be derived from the spectral Doppler wave forms accordingly. Figure 1(h) shows an example of the maximum velocity envelope derived from the spectral Doppler wave forms in Fig. 1(g). For each cardiac cycle in Fig. 1(h), symbol \( S \) represents the peak systolic maximum velocity, symbol \( D \) represents the end diastolic maximum velocity and symbol \( A \) represents the temporal average of maximum velocity. The PI and RI are defined as follows to characterize the curve and remove the dependence on Doppler angle:

\[
PI = \frac{(S-D)}{A}, \quad RI = \frac{(S-D)}{S}.
\]

Similar Doppler indices, such as the \( S/D \) ratio and \( D/A \) ratio, can be defined accordingly. Cycle 2 was excluded for statistical calculation due to eye motion. Table 1 summarizes the flow indices measured for other cardiac cycles, their average values, standard deviation values, and coefficients of variance for the maximum velocity envelope curve. The quantitative indices indicated above can be derived accordingly for other envelope curve definitions.

The intersession repeatability\(^{18}\) of RI measurement was assessed from multiple pairs of measurements. One measurement pair was defined as two separate measurements on the same vessel site. We measured two retinal vessels from two normal patients. Each vessel was measured seven times in the same day. There were \( 2 \times 7 \) independent measurement pairs out of seven repeated measurements on each vessel. The intersession coefficient of repeatability\(^{18}\) (CoR) of 0.08336 was calculated from the 42 pairs of measurements on two vessels (see Table 2) according to the formula: \( \text{CoR} = 1.96 \sqrt{\frac{\sum (d_0-d_1)^2}{n-1}} \).

In summary, we have developed a functional Fourier domain optical coherence tomography instrument that allows spectral Doppler flow imaging of \textit{in vivo} human retinal flow. An improved phase-resolved algorithm was developed to correct the bulk motion artifacts. The CoR was assessed for RI measurements using 14 measurements of two vessels within two normal subjects. This method provides an alternative way to quantify retinal blood flow with Doppler-angle-independent flow indices that may provide insight on the retinal flow in many vascular related eye diseases.

**Acknowledgment**

This work is supported by the National Institutes of Health (Grant No. EB-00293, No. NCI-91717, No. RR-01192), and the Air Force Office of Science Research (Grant No. FA9550-04-1-0101). Institutional support from the Beckman Laser Institute and Medical Clinic is also gratefully acknowledged.

**References**