

Journal of Biomedical Optics

SPIEDigitalLibrary.org/jbo

Laser speckle contrast imaging: theoretical and practical limitations

David Briers
Donald D. Duncan
Evan Hirst
Sean J. Kirkpatrick
Marcus Larsson
Wiendelt Steenbergen
Tomas Stromberg
Oliver B. Thompson

Laser speckle contrast imaging: theoretical and practical limitations

David Briers,^a Donald D. Duncan,^b Evan Hirst,^c Sean J. Kirkpatrick,^d Marcus Larsson,^e Wiendelt Steenbergen,^f Tomas Stromberg,^e and Oliver B. Thompson^c

^aEmeritus Professor, Kingston University, United Kingdom

^bPortland State University, Maseeh College of Electrical and Computer Science, P.O. Box 751, Portland, Oregon 97207

^cCallaghan Innovation, P.O. Box 31-310, Lower Hutt, 5040 New Zealand

^dMichigan Technological University, Department of Biomedical Engineering, M&M Building 301, 1400 Townsend Drive, Houghton, Michigan 49931

^eLinköping University, Department of Biomedical Engineering, SE-581 83 Linköping, Sweden

^fUniversity of Twente, MIRA Institute, PO Box 217, NL-7500 AE Enschede, The Netherlands

Abstract. When laser light illuminates a diffuse object, it produces a random interference effect known as a speckle pattern. If there is movement in the object, the speckles fluctuate in intensity. These fluctuations can provide information about the movement. A simple way of accessing this information is to image the speckle pattern with an exposure time longer than the shortest speckle fluctuation time scale—the fluctuations cause a blurring of the speckle, leading to a reduction in the local speckle contrast. Thus, velocity distributions are coded as speckle contrast variations. The same information can be obtained by using the Doppler effect, but producing a two-dimensional Doppler map requires either scanning of the laser beam or imaging with a high-speed camera: laser speckle contrast imaging (LSCI) avoids the need to scan and can be performed with a normal CCD- or CMOS-camera. LSCI is used primarily to map flow systems, especially blood flow. The development of LSCI is reviewed and its limitations and problems are investigated. © The Authors. Published by SPIE under a Creative Commons Attribution 3.0 Unported License.

Distribution or reproduction of this work in whole or in part requires full attribution of the original publication, including its DOI. [DOI: [10.1117/1.JBO.18.6.066018](https://doi.org/10.1117/1.JBO.18.6.066018)]

Keywords: laser speckle; laser Doppler; time-varying speckle; medical imaging; blood flow; perfusion.

Paper 130209R received Apr. 3, 2013; revised manuscript received May 17, 2013; accepted for publication May 17, 2013; published online Jun. 27, 2013.

1 Introduction

The first part of this paper is a review of the technique known variously as laser speckle contrast imaging (LSCI), laser speckle imaging (LSI), or laser speckle contrast analysis (LASCA). The technique uses the phenomenon of laser speckle. The basic theory of laser speckle was developed in the 1960s.¹ In the 1970s, time-varying speckle, caused by motion, became a subject for research. In particular, a connection was established between the fluctuations of the speckle pattern and the movement of scattering centers in living organisms, for example, the movement of red blood cells.² One way in which the speckle fluctuations manifest themselves is in a reduction in the normally high contrast of the speckle pattern. In the 1980s, this effect was used in a photographic technique known as single-exposure speckle photography, developed to study blood flow in the retina.³ Although the method worked, the need to process the photographs before the information could be accessed proved to be a major problem and interest in the technique waned. In the 1990s, new digital methods allowed the development of a real-time version of the method⁴ and this has proved to be much more useful. There are, however, some problems, both theoretical and practical, and the second part of the paper will attempt to address these.

2 Background

2.1 Laser Speckle

When laser light illuminates a diffuse surface, the high coherence of the light produces a random granular effect known as speckle. Figure 1 shows a typical speckle pattern.

Laser speckle is an interference pattern produced by light reflected or scattered from different parts of the illuminated surface. If the surface is rough (surface height variations larger than the wavelength of the laser light used), light from different parts of the surface within a resolution cell (the area just resolved by the optical system imaging the surface) traverses different optical path lengths to reach the image plane. (In the case of an observer looking at a laser-illuminated surface, the resolution cell is the resolution limit of the eye and the image plane is the retina.) The resulting intensity at a given point on the image is determined by the superposition of all waves arriving at that point. If the resultant amplitude is zero because all the individual waves cancel out, a dark speckle is seen at the point; if all the waves arrive at the point in phase, an intensity maximum is observed.

Laser speckle is a random phenomenon and can only be described statistically. Goodman¹ has developed a detailed theory, but for this paper only one result is of major importance. This is an expression for the contrast of a speckle pattern. Assuming ideal conditions for producing a speckle pattern—highly coherent, single-frequency laser light; linear polarization; and a perfectly diffusing surface—Goodman showed that the standard deviation of the intensity variations in the speckle

Address all correspondence to: David Briers, Cwm Gorllwyn, Tegryn, Llanfyrnach, SA35 0DN, United Kingdom. Tel: +44 1239 698264; E-mail David.Briers@physics.org

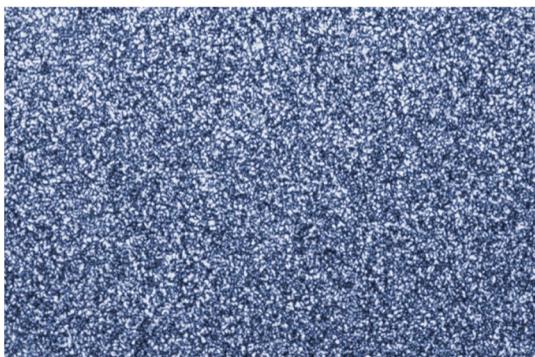


Fig. 1 A typical laser speckle pattern.

pattern is equal to the mean intensity. In practice, speckle patterns often have a standard deviation that is less than the mean intensity and this causes a reduction in the contrast of the speckle pattern. In fact, it is normal to define the speckle contrast as the ratio of the standard deviation to the mean intensity:

$$K = \frac{\sigma}{\langle I \rangle}. \quad (1)$$

Although a detailed account of laser speckle statistics is outside the scope of this paper, it is worth mentioning at this point that the scale of the speckle pattern—the size of the individual speckles—has, in general, nothing to do with the structure of the surface producing it. It is determined entirely by the wavelength of the light and the aperture of the optical system used to observe the speckle pattern. If the speckle pattern is being observed directly by the human eye, it is the pupil of the eye that determines the speckle size. More importantly, if a camera is used, it is the setting of the aperture stop that determines the speckle size. This can have a serious effect if the aperture is used to control the exposure of the image.

2.2 Time-Varying Speckle

When an object moves, the speckle pattern it produces changes. For small movements of a solid object, the speckle pattern moves as a whole, i.e., the speckles remain correlated. For larger motions, the speckles “decorrelate” and the speckle pattern changes completely. Decorrelation also occurs when the light is scattered from a large number of individual moving scatterers, such as particles in a fluid. An individual speckle appears to “twinkle” like a star. This phenomenon has come to be known as “time-varying speckle.” One of the most important potential applications of speckle fluctuations, first recognized by Stern,² arises when they are caused by the flow of blood.

It is reasonable to assume⁵ that the frequency spectrum of the fluctuations should be dependent on the velocity of the motion. It should therefore be possible to obtain information about the motion of the scatterers from a study of the temporal statistics of the speckle fluctuations. This is the basis of the study of time-varying speckle, many of whose applications have been in the biomedical field.

2.3 Relationship with Laser Doppler

Movement, especially of individual scatterers, causes laser speckle patterns to fluctuate in time. However, laser Doppler

techniques also analyze the frequency spectrum of light intensity fluctuations observed when laser light is scattered from moving particles. Are these the same fluctuations? The physics at first sight looks different in the two cases. In the Doppler method, the frequency of light scattered from moving particles is assumed to be frequency-shifted and this “beats” with non-shifted light from stationary parts of the object (or from a reference beam) to give a Doppler signal whose frequency is equal to the difference between the two frequencies. On the other hand, no frequency shift is invoked to explain time-varying speckle—the speckle pattern is produced by interference of light of the same frequency that has traversed different optical path lengths to reach the detector, and the fluctuations are caused by these path lengths changing as a result of the motion of the scatterers. However, the two techniques yield the same mathematical formula connecting the frequency of the fluctuations and the velocity of the scatterers⁶—they are simply two different ways of looking at the same phenomenon.

Whether regarded as Doppler or as time-varying speckle, it is important to note that measurements of the temporal statistics of the intensity fluctuations can, in principle, be carried out only at a single point (strictly, a single speckle). If a map of the velocity is required, some method of scanning is necessary. This has been done for both speckle^{7–10} and for Doppler.^{11–13} The main problem with these scanning instruments is the time taken for a scan to be carried out and for the data to be processed—typically several minutes. It was for this reason that the technique of LSCI, which produces a map of velocity in a single shot, was developed.

Some workers claim that the main difference between LSCI and laser Doppler is that the former is qualitative and the latter quantitative. In other words, LSCI needs to be calibrated and Doppler does not. We believe there is some confusion here and this needs to be addressed.

It is true that the Doppler technique, as originally envisaged, gives absolute measurements of velocity, but only in a small and well-defined volume, typically 0.1 mm³ or less, that is defined by two or more laser beams crossing at an oblique angle. These systems are capable of providing, without calibration, absolute velocities in one or more dimensions (depending on the number of laser beams used). One of the beams is typically frequency-shifted, which allows the direction of the movement to be determined and hence the full velocity vector to be measured.

Note that in this paper we are following the current practice of referring to “velocity” when we really mean just its magnitude; when the direction of travel is also known, we shall refer to the “full velocity vector,” as above. We are also using the clinical term “perfusion” for blood flow: the accepted units for this are typically milliliters per 100 grams per minute, or sometimes milliliters per 100 milliliters per minute. This clearly involves concentration and contrasts with other types of flow, where the units for “rate of flow” are volume per unit time. In this paper, we shall use “flow” to mean “rate of flow” and “perfusion” when we are talking specifically about blood flow. It is important, however, to remember the above differences in definition and units.

In contrast to the technique described earlier in this section, the Doppler systems used for perfusion measurements are “regional” rather than point-wise. They still rely on the Doppler shift, but in this case multiple scattering in static structures surrounding the blood vessels will blur the relationship

between the direction of the blood flow and the scattering vector. This is further pronounced in tissue, where blood flows in a variety of directions. As a result, a single blood flow velocity will give rise to a distribution of Doppler frequencies that depends not only on the blood flow velocity and concentration but also on the scattering phase function of the red blood cells and the degree of multiple Doppler shifts.

A measure of motion is derived by calculating the first-order moment of this Doppler spectrum, but such measurements provide only relative estimates of regional perfusion (not absolute measurements, as with point-wise systems). These regional measurements are of value when the objective is to measure relative changes in perfusion, for example, due to some stimulus. Quantitative assessment of volumetric flow with these systems is very error-prone and certainly requires calibration.

Note that the above discussion applies equally to both regional laser Doppler systems (such as those used for perfusion measurements) and LSCI. In their original forms, neither can produce absolute measurements of velocity and both require calibration. Fredriksson et al. have recently proposed a tissue and light transport modeling approach aimed at absolute perfusion estimation.¹⁴

We should mention at this point that there are other techniques for imaging blood flow, including spectroscopic methods such as tissue viability and hyperspectral imaging. Our intention in this paper is not to compare LSCI with these other techniques, and we would refer the reader to other publications for such comparisons.^{15,16}

3 History

3.1 Single-Exposure Speckle Photography

In the early 1980s, Fercher and Briers³ introduced the idea of using speckle contrast reduction to measure flow. They called the technique “single-exposure speckle photography,” in order to distinguish it from the double-exposure method widely used to measure simple movements.

The basic argument is that in a photograph taken under laser illumination, the speckle pattern in an area where flow is occurring is blurred to an extent that depends on the velocity of flow and on the exposure time of the photograph. The speckle pattern in an area of no flow, on the other hand, remains of high contrast. Thus velocity distributions are mapped as variations in speckle contrast.

In practice, contrast variations are difficult for the human eye to detect and some method of enhancing the contrast maps is necessary. Digital techniques were not sufficiently developed in the early 1980s for this to be done as the photograph was taken (though they could have been used on the resulting photograph). Fercher and Briers found, however, that a simple optical filtering process, using a high-pass spatial filter, worked quite well and resulted in the contrast variations being converted into intensity variations. They successfully applied the technique to the mapping of retinal blood flow.¹⁷ Figure 2 shows an example from their 1982 paper.

Although the feasibility of single-exposure speckle photography had been demonstrated, the fact that it was a two-step process—the photograph had first to be processed, the resulting transparency had to be placed in the spatial filtering setup, and then a second photograph had to be taken—reduced its attractiveness to clinicians and researchers.

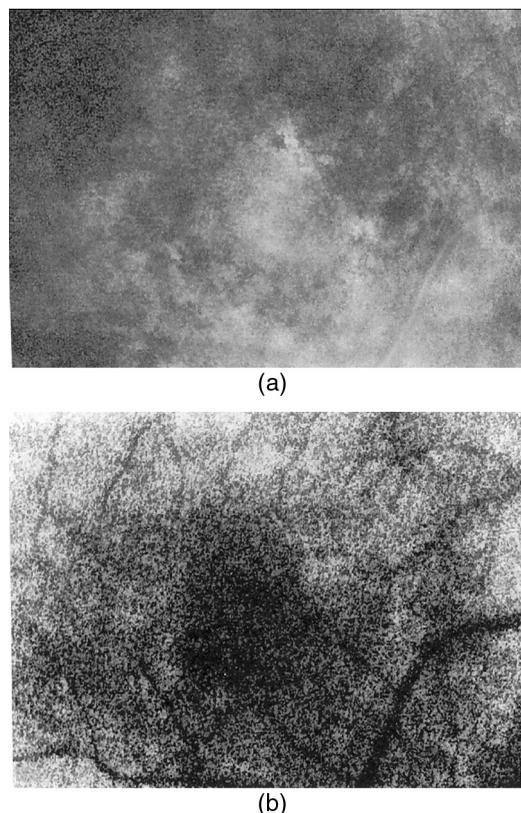


Fig. 2 Single-exposure speckle photography¹⁷—raw image of part of a retina (a) and its spatially filtered version, showing contrast variations mapped as intensity variations (b).

3.2 Laser Speckle Contrast Imaging: a Digital Version of Single-Exposure Speckle Photography

By 1990, digital techniques were sufficiently advanced to justify taking another look at single-exposure speckle photography. Briers and Webster⁴ succeeded in measuring the contrast directly and converting it to a false-color image, thus avoiding the main disadvantage of the photographic process. As the procedure no longer involved photography, a new name was needed, and they suggested LASCA. Today, alternative names include LSCI and LSI. Figure 3 shows an early example of the original LASCA technique.⁴

3.3 Some Recent Work on Laser Speckle Contrast Imaging

Several of the authors of this paper—and many others around the world—have developed and improved the techniques of LSCI over the past two decades. Examples include optimization of the exposure time by Boas’s group,¹⁸ a noise reduction scheme by Scheffold’s group,¹⁹ and some significant contributions to the theory.^{20–22}

Applications have been mainly in the medical field, as expected, with a lot of activity in using the technique to monitor cerebral blood flow.^{23–28} Boas’s group has been particularly active in this area and has also used the technique in an investigation into migraines.²⁹ Other medical applications have included microcirculation investigations,^{30–32} dentistry,³³ wound and burn assessment,^{34–36} and a return to ophthalmological problems.^{37–40} Nonmedical applications have included measuring the velocity of vehicles^{41,42} and monitoring the drying of paint.⁴³

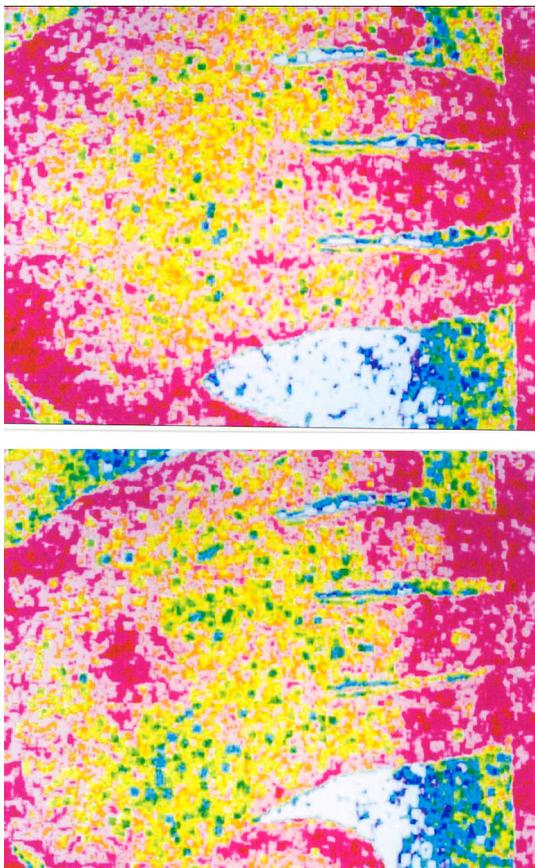


Fig. 3 LASCA images of the back of a hand,⁴ showing a change in perfusion caused by rubbing a small area: blue indicates high contrast and therefore little or no flow, while red indicates low contrast and therefore high flow.

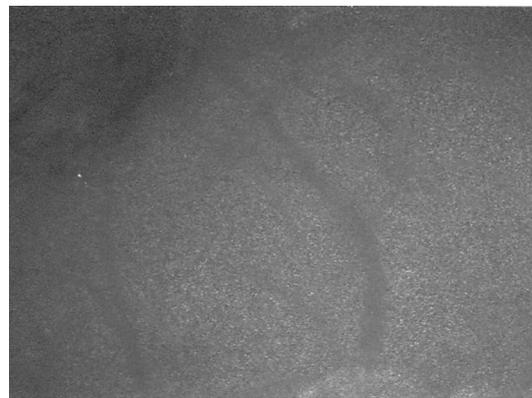
In addition to the above (and much other) work, there have been several reviews of speckle contrast imaging,^{44–48} including some comparative studies with laser Doppler techniques.^{49–51}

This recent work on LSCI has, of course, been accompanied by improvements in the images produced. Figures 4 and 5 are just two examples—the improvement in the quality of the images is clear (see Figs. 2 and 3).

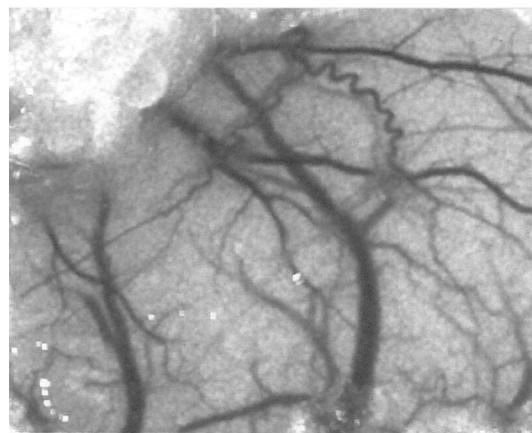
In recent years, at least two companies have launched instruments based on LSCI, both with real-time video capability. These allow the operator to follow changes of flow (in particular, blood perfusion) in real time.

4 Principle

The experimental setup for LSCI is very simple. Laser light illuminates the object under investigation, which is imaged by a digital camera. The image is captured and processed by custom software. The operator usually has several options at his disposal. In the original LASCA technique,⁴ this included the exposure time, the number of pixels over which the local contrast was computed, the scaling of the contrast map, and the choice of colors for coding the contrast. The choice of the number of pixels over which to compute the speckle contrast is important—too few pixels lead to the statistics being compromised and too many cause spatial resolution to be sacrificed.⁵² In practice, it is found that a square of 7×7 or 5×5 pixels is usually a satisfactory compromise. (A square with sides of an odd



(a)



(b)

Fig. 4 Raw image of part of a rat cortex (a) and its LSCI version (b).

number of pixels was chosen so that the computed contrast could be assigned to the central pixel.) The speckle contrast K is quantified by the usual parameter of the ratio of the standard deviation to the mean ($\sigma / \langle I \rangle$) of the intensities recorded for each pixel in the square [see Eq. (1)]. The pixel square is then moved along by 1 pixel and the calculation repeated: this overlapping of the pixel squares results in a much smoother image than would be obtained by using contiguous squares, and at little cost in terms of additional processing time. It must be remembered, though, that this overlapping of the squares does not lead to an increase in resolution, which is determined by the size of square used: there is a trade-off between spatial resolution and reliable statistics.

5 Theory

The original 1981 paper on single-exposure speckle photography by Fercher and Briers³ included a preliminary mathematical analysis. This made several rather bold assumptions about the statistics involved, but produced some promising results. The starting point was a formula first derived by Goodman⁵³ in 1965, connecting the variance of a time-averaged speckle pattern and the temporal statistics of the fluctuations. In 1985, Goodman⁵⁴ published a correction to his 1965 formula and the relationship between the variance of a time-averaged dynamic speckle pattern and the temporal fluctuation statistics is now given by

$$\sigma_s^2(T) = \frac{2}{T} \int_0^T \left(1 - \frac{\tau}{T}\right) C_t^{(2)}(\tau) d\tau, \quad (2)$$

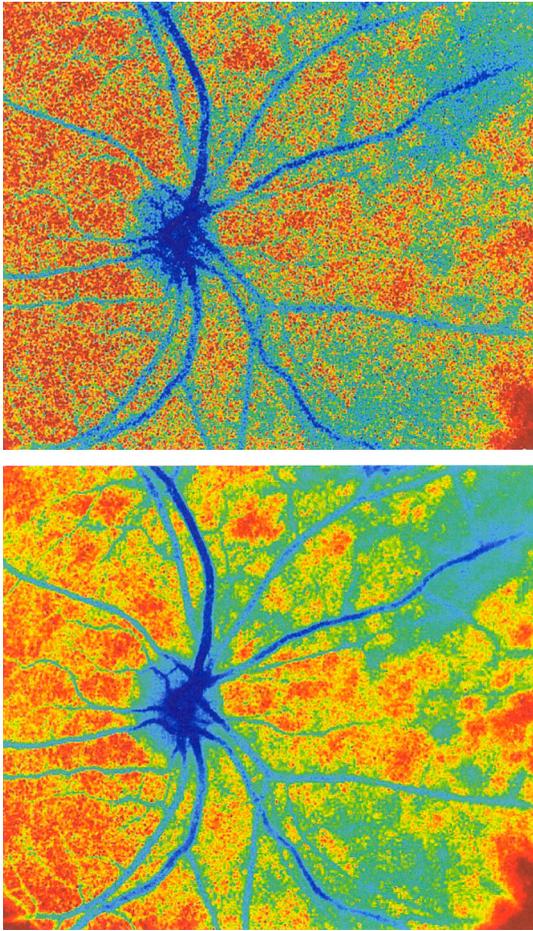


Fig. 5 LSCI images of part of a human retina, single exposure above and average of eight successive exposures below.

where $\sigma_s^2(T)$ is the variance of the spatial intensity distribution in a time-averaged speckle pattern with an exposure time (integration time) T and $C_t^{(2)}(\tau)$ is the autocovariance of the temporal fluctuations in the intensity fluctuations of a single speckle. $C_t^{(2)}(\tau)$ depends critically on the actual velocity distribution of the scattering particles and the proportion of the photons that are Doppler-shifted. Hence, to estimate the average velocity from a single-exposure image, both the fraction of Doppler-shifted photons and the velocity distribution must be either known or assumed.

Assuming all photons being Doppler-shifted and a Lorentzian velocity distribution, for example, leads to the following equation for the speckle contrast K as a function of the ratio of the correlation time to the exposure time (τ_c/T)

$$K = \frac{\sigma}{\langle I \rangle} = \left(\beta \left\{ \frac{\tau_c}{T} + \frac{\tau_c^2}{2T^2} \left[\exp\left(\frac{-2T}{\tau_c}\right) - 1 \right] \right\} \right)^{\frac{1}{2}}, \quad (3)$$

where β is an instrumentation-dependent constant introduced to account for the loss of correlation related to the ratio of the detector (or pixel) size to the speckle size, and to polarization.⁵⁵ The correlation time τ_c is the time taken for the contrast to fall to a specific level. It is inversely proportional to the local velocity of the scatterers. The above function is plotted as the curve labeled Lorentzian in Fig. 6. The speckle contrast rises from

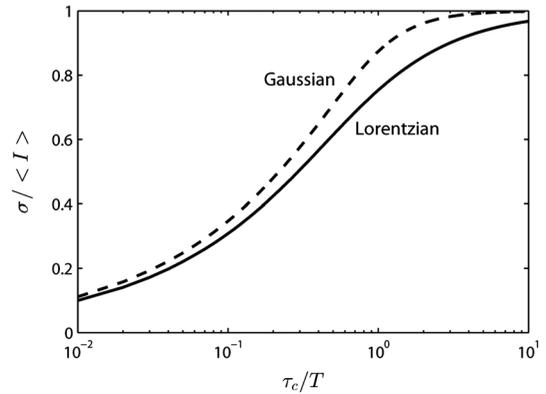


Fig. 6 Theoretical relationship between speckle contrast and the ratio of the speckle correlation time to exposure time, assuming a Lorentzian and a Gaussian velocity distribution, respectively. (Note that β has been set to 1 and the two curves have been normalized so that they can be compared.)

near zero to near its maximum value of 1.0 over about two orders of magnitude of τ_c (and hence of velocity). (For a single exposure, of course, T is a constant.) For velocities corresponding to values of τ_c less than about $0.04T$, the speckle contrast is very low, i.e., the speckles are completely blurred out by the motion. For velocities corresponding to values of τ_c greater than about $4T$, the speckle pattern remains almost fully developed, with maximum contrast. Between these limits, the velocity distribution is mapped as a variation in speckle contrast.

The curve for a Gaussian velocity distribution is also plotted in Fig. 6. It, too, shows the characteristic S-shape, but with a steeper slope. In addition, the curves have been normalized in order to compare them—they do not naturally fall in the same range of τ_c/T . It is clear, therefore, that the actual relationship between speckle contrast and τ_c/T (and hence the measured velocity) depends critically on the velocity distribution.

In principle, Eq. (3) provides the link between speckle contrast and velocity. However, the equation has been derived by making several assumptions and approximations, some of them being quite drastic. In particular, a Lorentzian velocity distribution has been assumed. Changing the shape of the velocity distribution will significantly affect the shape of the curve shown in Fig. 6, and hence the relationship between speckle contrast and velocity. This is just one of the several problems that we shall address in the next part of this paper.

6 Problems

6.1 Velocity Distribution

Equation (2) shows the relationship between the normalized variance of a time-integrated fluctuating speckle pattern (speckle contrast) and the temporal statistics of the fluctuations (autocovariance). LSCI measures the quantity on the left-hand side of this equation. Laser Doppler, on the other hand, directly measures the temporal statistics of the fluctuations (provided the concentration of moving scatterers is not too large), effectively measuring $C_t^{(2)}(\tau)$ in the right-hand side of the equation. For tissues containing a low concentration of red blood cells, it is widely accepted that the first moment of the Doppler spectrum (the power spectrum of the fluctuations) scales linearly with velocity and concentration.⁵ This means that the regional Doppler techniques used for blood perfusion measurements

(see Sec. 2.3) measure changes in the tissue perfusion. In the case of single-exposure LSCI, however, the link between the spatial statistics (speckle contrast) and tissue perfusion can only be made if the velocity distribution is known. In general, this will not be the case. Equation (3), linking speckle contrast with velocity, has been derived by assuming a particular form for the velocity distribution. It is clear from Fig. 6 and the discussion of it above that the choice of velocity distribution has a major effect on this relationship.

The original work on single-exposure speckle interferometry³ and LASCA⁴ assumed a Lorentzian distribution. This is probably appropriate for Brownian motion (unordered flow), but for ordered flow, a Gaussian distribution is usually considered more appropriate. As Duncan and Kirkpatrick have pointed out, there is an argument that the actual distribution is some combination of the two.⁵² It is clear from Fig. 6 that a measurement using a single integration time (and hence a single value of τ_c/T) cannot determine which velocity distribution curve is the correct one to use. The question remains as to whether the actual velocity distribution can be determined by other methods and then used to quantify LSCI measurements.

A related issue arises from the fact that LSCI computes the speckle contrast at each point by using the local standard deviation and local mean intensity. This has its own probability distribution, which Duncan and Kirkpatrick have shown to be log-normal.⁵⁶ The result of this is that any velocity estimate derived on the basis of computed local speckle contrast will be a sample statistic with its own attendant probability distribution.

Strictly speaking, the speckle contrast as measured by LSCI is dependent on the correlation time, τ_c , and it is usually accepted that this is inversely proportional to some “typical” velocity. However, the constant of proportionality is open to question and depends to some extent on the direction of motion of the scatterers. This will clearly have some impact on the ability of LSCI to measure absolute velocities, flow, or perfusion. A related question is whether non-Newtonian flow (which blood perfusion certainly is) could be an issue.

6.2 Velocity or Flow?

Equation (3) relates speckle contrast to the correlation time τ_c , which is inversely proportional to velocity. It is widely accepted, however, that laser Doppler measures flow (perfusion in the case of blood flow). The question arises: does LSCI measure velocity or flow?

It is easy to show that the speckle contrast must be affected by the number of moving scatterers involved, and hence by the concentration, as this affects the fraction of Doppler-shifted photons. A speckle pattern produced only by stationary scatterers under ideal conditions, will have a speckle contrast of 1 (the maximum). If just a few moving particles are added, it is clear that some intensity fluctuations will be introduced, so that a time-integrated image of the speckle pattern will show some loss of contrast. However, the intensity of each individual speckle will show only small fluctuations about its original (stationary) value. This means that the speckle pattern will be dominated by the pattern from the stationary scatterers and the loss of contrast will be small, even for long integration times. As the number of moving scatterers is increased, the effect of the stationary scatterers will diminish and, for a given integration time, the speckle contrast will decrease. The effect on the graph of speckle contrast against τ_c/T is illustrated

schematically in Fig. 7. It is clear that a measurement using a single integration time (and hence a single value of τ_c/T) cannot determine whether the continuous or the broken (or any other) curve is the correct one to use.

In 1978, Briers⁵⁷ presented a theoretical analysis of the speckle contrast produced by a mixture of moving and stationary scatterers over a long integration time and deduced the following simple relationship between K , the speckle contrast, and ρ , the fraction of photons in the scattered light that are Doppler-shifted:

$$K = 1 - \rho. \quad (4)$$

In 2003, Rabal et al.⁵⁸ confirmed this equation experimentally. In theory, Eq. (4) could be used on a long-exposure LSCI image to fix the minimum-contrast point on the broken curve of Fig. 7, a contrast value that is strongly dependent on the concentration of moving scatterers rather than their velocity. However, it should be noted that the presence of a static component may also change the shape of the curve.^{59,60} From Figs. 6 and 7, it is clear that single-exposure LSCI cannot be related to perfusion in the same way as laser Doppler, without knowledge or assumptions regarding the velocity distribution and the fraction of photons that are Doppler-shifted.

6.3 Multiple Scattering

It is usually assumed that the photons detected in LSCI have been scattered only once from a moving blood cell. However, it is becoming increasingly clear that some multiple scattering will occur. In tissues with high blood volume fractions, such as the brain, even if light scatters within a vessel no more than once, there is a high probability of detected photons scattering from more than one blood cell from different vessels. One effect of this will be that the technique will be sensitive to the relative motion of blood cells as well as to their absolute motion. As discussed in Sec. 2.3, multiple scattering also means that even a single blood flow velocity will give rise to a distribution of Doppler frequencies, i.e., a spectrum. This leads to the need for both LSCI and Doppler systems, when used to monitor perfusion, to be calibrated.

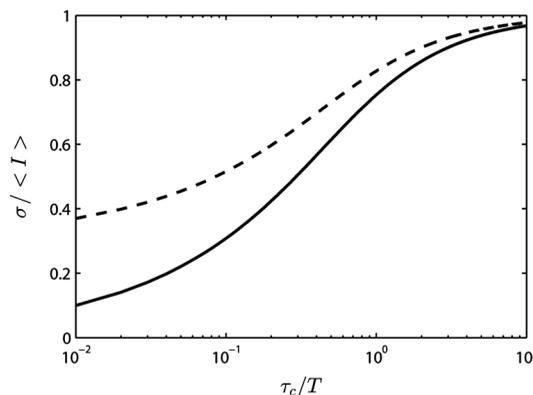


Fig. 7 Theoretical speckle contrast as a function of the ratio of the speckle correlation time to exposure time, for a completely dynamic medium (solid line) and a medium with a fraction of stationary scatterers (broken line) (schematic only).

6.4 Speckle Size and Polarization

In Eq. (3), the factor β is intended to account for the loss of correlation related to the ratio of the detector size to the speckle size, and to polarization.⁵⁵ However, it is not clear whether the invoking of this factor can accurately and reliably compensate for these problems. Kirkpatrick et al.⁶¹ have carried out a detailed investigation of the effect of speckle size on speckle contrast. Thompson et al.²² have shown that a linear β correction is valid for simple phantoms and this result may be generalizable to tissue measurements.

7 Analysis

7.1 Velocity Distribution

In order to make the link between the spatial statistics of single-exposure time-integrated speckle patterns (used in LSCI) and the temporal statistics of the intensity fluctuations (used in laser Doppler), it is necessary to know, or assume, the form of the velocity distribution. Poor assumptions are, however, likely to introduce significant errors. It is possible, though, that initial experiments on the type of target to be used might lead to an improved approximation for the distribution, which could then be used in Eq. (3). In 2008, Duncan and Kirkpatrick⁵² suggested a more physically realistic description of the velocity distribution, based on the normalized intensity covariance as expressed by Goodman¹. This “rigid-body” model is an intermediate between the limiting Lorentzian and Gaussian solutions and closely resembles the Lorentzian expectations for long exposures (relative to the correlation time) and the Gaussian expectations for short exposures.

The alternative is to measure the velocity distribution. Thompson and Andrews⁶² have suggested that this can effectively be done by using multiple exposures. They showed that between 10 and 15 exposures, with each successive exposure being double the previous one, are sufficient to produce data on a par with Doppler techniques. The question that arises is whether this can be done while maintaining the real-time advantage of LSCI.

7.2 Velocity or Flow?

For laser Doppler methods, there is a generally accepted link between the first-order moment of the power spectrum and flow (perfusion in the case of blood).⁵ There is no such accepted theory in the case of laser speckle contrast techniques. Figure 7 shows qualitatively that the presence of stationary (or very slow-moving) scatterers in the field affects the measured speckle contrast. Hence, flow, which depends on the fraction of moving scatterers (e.g., blood in tissue), must have an effect. Whether the technique measures flow, or some quantity related to flow, is an open question and merits further work. Some work has been done on how the presence of static background speckle (e.g., from nonmoving tissue) affects LSCI measurements and some success has been achieved, notably by Zakharov et al.⁵⁹ and by Parthasarathy et al.⁶³ Another possibility might be to combine LSCI with other concepts, such as structured illumination, as suggested by Cuccia et al.⁶⁴

Some initial work by Draijer et al.⁶⁵ has shown that using the parameter $1 - K^2$ rather than K has some advantages, in that it can be shown to be a frequency-weighted integral of the power spectrum. In fact, as the integration time T goes to infinity, the quantity $1 - K^2 \rightarrow M_0$, the zero-order moment of the power

spectrum, and hence depends strongly on the concentration of moving particles. This to some extent quantifies Fig. 7, as $T \rightarrow \infty$ implies $\tau_c/T \rightarrow 0$. It is worth investigating whether $1 - K^2$ gives a better agreement with laser Doppler measurements. However, with the inclusion of non-Doppler-shifted photons, Eq. (3) needs to be revised.

7.3 Correlation Time and Velocity

Further work is needed on the actual relationship between correlation time τ_c and velocity. Formulae in the literature vary by factors of up to 30 or more, depending on the assumptions made (especially on the effect of multiple scattering). Simple dimensional arguments require that velocity and correlation time be related through a spatial scale length. This is usually taken to be the wavelength of the light, but it is the dimensionless multiplying constant that causes the problems. Perhaps we can avoid the problems if we can relate the speckle measurements to flow rather than velocity.

7.4 Multiple Doppler Scattering

There is no doubt that multiple Doppler scattering is highly likely to occur and may be difficult to quantify. The degree of multiple scattering will depend on the circumstances of the field being monitored and may well be different for each measurement made. A theoretical solution to this problem may well be insoluble, although Monte Carlo techniques may go some way toward this.

There is also a potentially very significant issue in the presence of multiple populations of scatterers with different correlation times (and hence velocities), all within the same depth of field. The scattered light from the different populations will combine to give a time-varying speckle pattern with a decorrelation behavior somewhere between those of the two (or more) populations independently. It is possible that blind deconvolution techniques may find a solution, but the lack of *a priori* information about multiple populations will make this very difficult, and perhaps also insoluble. Note that both these problems may also affect laser Doppler measurements.

7.5 Speckle Size and Polarization

Both these factors will affect the absolute interpretation of the speckle contrast in terms of flow or velocity, but it is less clear whether or not they will affect relative measurements. Hence, although further work is needed on their impact, it is possible that they will not affect measurements of changes in flow (or perfusion in the case of blood), or of variations in flow across an image.

8 Conclusions and Recommendations

8.1 Calibration

Although there is no doubt that LSCI is a powerful technique for mapping blood perfusion (and other flow fields) in real time, the physics of the scattering process is so complex and indeterminate that we believe it might never be possible to make absolute measurements. Work will no doubt continue on trying to find solutions to the problems, but in the meantime our recommendation is to regard LSCI as a semi-quantitative technique that requires calibration. (Note that the discussion in Sec. 2.3

indicates that regional Doppler techniques, such as those used in perfusion measurement systems, also require calibration.)

Users of the technique (including the manufacturers of commercial instruments) tend to use a calibration on a phantom to fix a point on a scale and all measurements are made relative to this in arbitrary “perfusion units.” The speckle contrast values are not converted to absolute values of flow, nor are they linear relative to absolute flow.

Because of the uncertainty surrounding the actual velocity distribution that should be used, one approach is to use a much simpler, arbitrary function that produces an S-shaped curve similar to that of Fig. 6. This is done in order to simplify the algorithm and speed up the processing. Possible candidates include the functions $1/K - 1$ and $1/K^2 - 1$ (or their square roots). It can be seen that both these functions go to zero as K approaches 1 and go to infinity as K approaches 0, as required. They are, of course, arbitrary, but the argument is that any velocity distribution chosen is also arbitrary.

8.2 Multiple Exposures

Thompson and Andrews⁶² have shown that the velocity distribution problem might be solved by using multiple exposures with different integration times. This allows the Doppler spectrum to be computed. If this can be done quickly enough to preserve the real-time operation of LSCI, then it could go a long way toward solving one of the key problems of the technique. It may also answer the velocity/flow argument, though this is not yet clear. Two of the key questions to be answered are the number of exposures required and whether the technique can be made robust against motion artifacts. We believe this approach should be investigated further and that manufacturers should consider incorporating a multiple-exposure option into their instruments.

8.3 Future

The present situation is that LSCI is a valuable technique for the semi-quantitative real-time mapping of flow fields (including blood perfusion), but that it has to be calibrated and the results are in arbitrary units and not directly related to (or linear with) actual flow values. (Note that the Doppler technique also requires calibration, as discussed in Sec. 2.3.)

The incorporation of multiple exposures will, we believe, improve the quantification of LSCI by effectively allowing the velocity distribution and the fraction of photons that are Doppler-shifted to be measured. The number of exposures needed will have to be investigated.

Further theoretical work, including techniques such as Monte Carlo simulations and blind deconvolution, may improve the robustness of the theory, but not, we think, to the extent that a truly quantitative technique can be achieved.

Because of the complexity of the physical processes and the need (at present) to calibrate LSCI instruments, we recommend that some effort be put into the formulation of a standard experimental configuration for LSCI experiments. (Again, note that the same arguments apply to Doppler techniques.)

In the long term, it is likely that improvements in computer power will allow the parallel processing of laser Doppler images to produce real-time maps. In fact, initial steps to realize this have already been made.⁶⁶ We believe, however, that LSCI will still offer some advantages, for example, where the more expensive Doppler techniques would be an overkill or when

maximum temporal resolution is required, and that it will continue to be a valuable tool for the real-time mapping of blood perfusion and other flow fields.

Acknowledgments

We are grateful to David A. Boas of the Harvard Medical School for his valuable contributions in the early stages of this project and for providing Fig. 4.

References

1. J. W. Goodman, “Statistical properties of laser speckle patterns,” in *Laser Speckle and Related Phenomena*, J. C. Dainty, Ed., Vol. 9 in series Topics in Applied Physics, pp. 9–75, Springer, Berlin, Heidelberg (1975).
2. M. D. Stern, “*In vivo* evaluation of microcirculation by coherent light scattering,” *Nature* **254**, 50–58 (1975).
3. A. F. Fercher and J. D. Briers, “Flow visualization by means of single-exposure speckle photography,” *Opt. Commun.* **37**(5), 326–330 (1981).
4. J. D. Briers and S. Webster, “Laser speckle contrast analysis (LSCA): a non-scanning, full-field technique for monitoring capillary blood flow,” *J. Biomed. Opt.* **1**(2), 174–179 (1996).
5. R. Bonner and R. Nossal, “Model for laser Doppler measurements of blood-flow in tissue,” *Appl. Opt.* **20**(12), 2097–2107 (1981).
6. J. D. Briers, “Laser Doppler and time-varying speckle: a reconciliation,” *J. Opt. Soc. Am. A* **13**(2), 345–350 (1996).
7. H. Fujii et al., “Evaluation of blood flow by laser speckle image sensing: part 1,” *Appl. Opt.* **26**(24), 5321–5325 (1987).
8. H. Fujii, “Visualization of retinal blood flow by laser speckle flowgraphy,” *Med. Biol. Eng. Comput.* **32**(3), 302–304 (1994).
9. Y. Tamaki et al., “Non-contact, two-dimensional measurement of retinal microcirculation using laser speckle phenomenon,” *Inv. Ophthalmol. Vis. Sci.* **35**(11), 3825–3834 (1994).
10. N. Konishi and H. Fujii, “Real-time visualization of retinal microcirculation by laser flowgraphy,” *Opt. Eng.* **34**(3), 753–757 (1995).
11. T. J. H. Essex and P. O. Byrne, “A laser Doppler scanner for imaging blood flow in skin,” *J. Biomed. Eng.* **13**(3), 189–194 (1991).
12. K. Wårdell, A. Jakobsson, and G. E. Nilsson, “Laser Doppler perfusion imaging by dynamic light scattering,” *IEEE Trans. Biomed. Eng.* **40**(4), 309–316 (1993).
13. K. Forrester, M. Doschak, and R. Bray, “*In vivo* comparison of scanning technique and wavelength in laser Doppler perfusion imaging: measurement in knee ligaments in adult rabbits,” *Med. Biol. Eng. Comput.* **35**(6), 581–586 (1997).
14. I. Fredriksson, M. Larsson, and T. Strömberg, “Model-based quantitative laser Doppler flowmetry in skin,” *J. Biomed. Opt.* **15**(5), 057002 (2010).
15. J. O’Doherty et al., “Comparison of instruments for investigation of microcirculatory blood flow and red blood cell concentration,” *J. Biomed. Opt.* **14**(3), 034025 (2009).
16. E. M. C. Hillman, “Optical brain imaging *in vivo*: techniques and applications from animal to man,” *J. Biomed. Opt.* **12**(5), 051402 (2007).
17. J. D. Briers and A. F. Fercher, “Retinal blood-flow visualization by means of laser speckle photography,” *Inv. Ophthalmol. Vis. Sci.* **22**(2), 255–259 (1982).
18. S. Yuan et al., “Determination of optimal exposure time for imaging of blood flow changes with laser speckle contrast imaging,” *Appl. Opt.* **44**(10), 1823–1830 (2005).
19. A. C. Völker et al., “Laser speckle imaging with an active noise reduction scheme,” *Opt. Express* **13**(24), 9782–9787 (2005).
20. A. Serov, W. Steenbergen, and F. de Mul, “Prediction of the photodetector signal generated by Doppler-induced speckle fluctuations: theory and some validations,” *J. Opt. Soc. A* **18**(3), 622–630 (2001).
21. R. Bandyopadhyay et al., “Speckle-visibility spectroscopy: a tool to study time-varying dynamics,” *Rev. Sci. Instrum.* **76**, 093110 (2005).
22. O. Thompson, M. Andrews, and E. Hirst, “Correction for spatial averaging in laser speckle contrast analysis,” *Biomed. Opt. Express* **2**(4), 1021–1029 (2011).
23. A. K. Dunn et al., “Dynamic imaging of cerebral blood flow using laser speckle,” *J. Cereb. Blood Flow Metab.* **21**(3), 195–201 (2001).

24. T. Durduran et al., "Spatiotemporal quantification of cerebral blood flow during functional activation in rat somatosensory cortex using laser-speckle flowmetry," *J. Cereb. Blood Flow Metab.* **24**(5), 518–525 (2004).
25. H. K. Shin et al., "Vasoconstrictive neurovascular coupling during focal ischemic depolarizations," *J. Cereb. Blood Flow Metab.* **25**(1), 1–13 (2005).
26. A. K. Dunn et al., "Spatial extent of oxygen metabolism and hemodynamic changes during functional activation of the rat somatosensory cortex," *Neuroimage* **27**(2), 279–290 (2005).
27. J. S. Paul et al., "Imaging the development of an ischemic core following photochemically induced cortical infarction in rats using laser speckle contrast analysis (LASCA)," *Neuroimage* **29**(1), 38–45 (2006).
28. P. Zakharov et al., "Dynamic laser speckle imaging of cerebral blood flow," *Opt. Express* **17**(16), 13904–13917 (2009).
29. H. Bolay et al., "Intrinsic brain activity triggers trigeminal meningeal afferents in a migraine model," *Nat. Med.* **8**(2), 136–142 (2002).
30. H. Cheng et al., "Laser speckle imaging of blood flow in microcirculation," *Phys. Med. Biol.* **49**(7), 1347–1357 (2004).
31. M. M. Gonik, A. B. Mishkin, and D. A. Zimnyakov, "Visualization of blood microcirculation parameters in human tissues by time-integrated dynamic speckles analysis," *Ann. NY Acad. Sci.* **972**, 325 (2002).
32. F. Domoki et al., "Evaluation of laser-speckle contrast image analysis techniques in the cortical microcirculation of piglets," *Microvasc. Res.* **83**(3), 311–317 (2012).
33. C. Stoianovici, P. Wilder-Smith, and B. Choi, "Assessment of pupal vitality using laser speckle imaging," *Lasers Surg. Med.* **43**(8), 833–837 (2011).
34. C. J. Stewart et al., "A comparison of two laser-based methods for determination of burn scar perfusion: laser Doppler versus laser speckle imaging," *Burns* **31**(6), 744–752 (2005).
35. C. J. Stewart et al., "Kinetics of blood flow during healing of excisional full-thickness skin wounds in pigs as monitored by laser speckle perfusion imaging," *Skin Res. Technol.* **12**(4), 247–253 (2006).
36. J. Qin et al., "Hemodynamic and morphological vasculature response to a burn monitored using a combined dual-wavelength laser speckle and optical microangiography imaging system," *Biomed. Opt. Express* **3**(3), 455–466 (2012).
37. M. Nagahara et al., "The acute effects of stellate ganglion block on circulation in human ocular fundus," *Acta Ophthalmol. Scand.* **79**(1), 45–48 (2001).
38. J. Flammer et al., "The impact of ocular blood flow in glaucoma," *Prog. Retin. Eye Res.* **21**(4), 359–393 (2002).
39. A. I. Srienec, Z. L. Kurth-Nelson, and E. A. Newman, "Imaging retinal blood flow with laser speckle flowmetry," *Front Neuroenergetics* **2**, 128 (2010).
40. W. Zhang et al., "Use of the laser speckle flowgraphy in posterior fundus circulation research," *Chin. Med. J. (Engl)* **124**(24), 4339–4344 (2011).
41. A. Aliverdiev, M. Caponero, and C. Moriconi, "Speckle velocimeter for a self-powered vehicle," *Tech. Phys.* **47**(8), 1044–1048 (2002).
42. D. Francis et al., "Objective speckle velocimetry for autonomous vehicle odometer," *Appl. Opt.* **51**(16), 3478–3490 (2012).
43. G. G. Romero, E. E. Alanis, and H. J. Rabal, "Statistics of the dynamic speckle produced by a rotating diffuser and its application to the assessment of paint drying," *Opt. Eng.* **39**(6), 1652–1658 (2000).
44. J. D. Briers, "Laser Doppler, speckle and related techniques for blood perfusion mapping and imaging," *Physiol. Meas.* **22**(4), R35–R66 (2001).
45. J. D. Briers, "Laser speckle contrast imaging for measuring blood flow," *Optica Applicata* **37**(1–2), 139–152 (2007).
46. M. Draijer et al., "Review of laser speckle contrast techniques for visualizing tissue perfusion," *Lasers Med. Sci.* **24**(4), 639–651 (2009).
47. D. A. Boas and A. K. Dunn, "Laser speckle contrast imaging in biomedical optics," *J. Biomed. Opt.* **15**(1), 011109 (2010).
48. K. Basak, M. Manjunatha, and P. K. Dutta, "Review of laser speckle-based analysis in medical imaging," *Med. Biol. Eng. Comput.* **50**(6), 547–558 (2012).
49. K. R. Forrester et al., "Comparison of laser speckle and laser Doppler perfusion imaging: measurement in human skin and rabbit articular tissue," *Med. Biol. Eng. Comput.* **40**(6), 687–697 (2002).
50. C. Millet et al., "Comparison between laser speckle contrast imaging and laser Doppler imaging to assess skin blood flow in humans," *Microvasc. Res.* **82**(2), 147–151 (2011).
51. G. A. Tew et al., "Comparison of laser speckle contrast imaging with laser Doppler for assessing microvascular function," *Microvasc. Res.* **82**(3), 326–332 (2011).
52. D. D. Duncan and S. J. Kirkpatrick, "Can laser speckle flowmetry be made a quantitative tool?," *J. Opt. Soc. Am. A* **25**(8), 2088–2094 (2008).
53. J. W. Goodman, "Some effects of target-induced scintillations on optical radar performance," *Proc. IEEE* **53**(11), 1688–1700 (1965).
54. J. W. Goodman, *Statistical Optics*, Wiley & Sons, New York (1985).
55. P. A. Lemieux and D. J. Durian, "Investigating non-Gaussian scattering processes by using nth-order intensity correlation functions," *J. Opt. Soc. Am. A* **16**(7), 1651–1654 (1999).
56. D. D. Duncan, S. J. Kirkpatrick, and R. K. Wang, "Statistics of local speckle contrast," *J. Opt. Soc. Am. A* **25**(1), 9–15 (2008).
57. J. D. Briers, "Statistics of fluctuation speckle patterns produced by a mixture of moving and stationary scatterers," *Opt. Quant. Electron.* **10**(4), 364–366 (1978).
58. H. J. Rabal et al., "Numerical model for dynamic speckle: an approach using the movement of the scatterers," *J. Opt. A: Pure & Appl. Opt.* **5**(5), S381–S385 (2003).
59. P. Zakharov et al., "Quantitative modelling of laser speckle imaging," *Opt. Lett.* **31**(23), 3465–3467 (2006).
60. P. Zakharov and F. Scheffold, "Advances in dynamic light scattering techniques," in *Light Scattering Reviews 4*, A. A. Kokhanovsky, Ed., pp. 433–467, Springer, Heidelberg (2009).
61. S. J. Kirkpatrick, D. D. Duncan, and E. M. Wells-Gray, "Detrimental effects of speckle-pixel size matching in laser speckle contrast imaging," *Opt. Lett.* **33**(24), 2886–2888 (2008).
62. O. B. Thompson and M. K. Andrews, "Tissue perfusion measurements: multiple-exposure laser speckle analysis generates laser Doppler-like spectra," *J. Biomed. Opt.* **15**(2), 027015 (2010).
63. A. B. Parthasarathy et al., "Robust flow measurement with multi-exposure speckle imaging," *Opt. Express* **16**(3), 1975–1989 (2008).
64. D. J. Cuccia et al., "Quantitation and mapping of tissue optical properties using modulated imaging," *J. Biomed. Opt.* **14**(2), 024012 (2009).
65. M. J. Draijer et al., "Relation between the contrast in time integrated dynamic speckle patterns and the power spectral density of their temporal intensity fluctuations," *Opt. Express* **18**(2), 21883–21891 (2010).
66. M. Draijer et al., "Twente optical perfusion camera: system overview and performance for video rate laser Doppler perfusion imaging," *Opt. Express* **17**(5), 3211–3225 (2009).