

Nondestructive assessment of the severity of occlusal caries lesions with near-infrared imaging at 1310 nm

Chulsung Lee
Dustin Lee
Cynthia L. Darling
Daniel Fried

University of California, San Francisco
Department of Preventive and Restorative Dental
Sciences
San Francisco, California 94143-0758

Abstract. The high transparency of dental enamel in the near-infrared (NIR) at 1310 nm can be exploited for imaging dental caries without the use of ionizing radiation. The objective of this study is to determine whether the lesion contrast derived from NIR imaging in both transmission and reflectance can be used to estimate lesion severity. Two NIR imaging detector technologies are investigated: a new Ge-enhanced complementary metal-oxide-semiconductor (CMOS)-based NIR imaging camera, and an InGaAs focal plane array (FPA). Natural occlusal caries lesions are imaged with both cameras at 1310 nm, and the image contrast between sound and carious regions is calculated. After NIR imaging, teeth are sectioned and examined using polarized light microscopy (PLM) and transverse microradiography (TMR) to determine lesion severity. Lesions are then classified into four categories according to lesion severity. Lesion contrast increases significantly with lesion severity for both cameras ($p < 0.05$). The Ge-enhanced CMOS camera equipped with the larger array and smaller pixels yields higher contrast values compared with the smaller InGaAs FPA ($p < 0.01$). Results demonstrate that NIR lesion contrast can be used to estimate lesion severity. © 2010 Society of Photo-Optical Instrumentation Engineers. [DOI: 10.1117/1.3475959]

Keywords: near-infrared imaging; occlusal surfaces; dental caries; transillumination.

Paper 10105PR received Mar. 1, 2010; revised manuscript received Jun. 4, 2010; accepted for publication Jun. 23, 2010; published online Aug. 6, 2010.

1 Introduction

Optical transillumination was used extensively before the discovery of x-rays for the detection of dental caries. Over the past two decades, there has been continued interest in this method, especially with the availability of high intensity fiber-optic-based illumination systems for the detection of interproximal lesions.¹⁻⁷ During fiber optic transillumination, a carious lesion appears dark because of decreased transmission due to increased scattering and absorption by the lesion. Several studies have been carried out using visible light transillumination, either as an adjunct to bitewing radiography or as a competing method for the detection of interproximal caries lesions.⁸⁻¹² However, since light scattering by sound enamel is significant in the visible range, the results are mixed.^{8,9}

Light scattering in sound enamel decreases markedly from the visible to the NIR.^{13,14} Even though light scattering for sound enamel is at a minimum near 1300 nm, the light scattering coefficient of enamel increases by 2 to 3 orders of magnitude on demineralization due to the formation of pores—on a similar size scale to the wavelength of the light—that act as Mie scatterers.¹⁵ Transmission measurements

through demineralized tissue sections at 1310 nm show that demineralized enamel attenuates NIR light by a factor 20 to 50 times greater than for sound enamel.^{16,17} Transillumination imaging at longer wavelengths in the NIR have been investigated in the region accessible to conventional silicon-based charge-coupled device (CCD) cameras, namely 830 nm^{18,19} and at 1310 to 1550 nm,^{16,17} where maximal transmission occurs. NIR images were acquired of natural interproximal and occlusal lesions through the transillumination of extracted human teeth.^{16,17}

Early enamel white spot lesions can be discriminated from sound enamel by visual observation or by visible-light diffuse reflectance imaging.^{20,21} The visibility of scattering structures on highly reflective surfaces such as teeth can be enhanced by use of crossed polarizers to remove the glare from the surface due to the strong specular reflection from the enamel surface.^{22,23} The contrast between sound and demineralized enamel can be further enhanced by depolarization of the scattered light in the area of demineralized enamel.^{24,25} The contrast between sound and demineralized enamel is greatest in the NIR due to the minimal scattering of sound enamel, and this can be exploited for reflectance imaging of early demineralization.¹⁵ Wu and Fried²⁶ reported the first high contrast polarized reflectance images of early demineralization on

Address all correspondence to: Daniel Fried, University of California, San Francisco, San Francisco, CA 94143-0758. Tel: 415-502-6641; Fax: 415-476-0858; E-mail: daniel.fried@ucsf.edu

buccal and occlusal tooth surfaces measured at $\lambda=1310$ nm, and found that the contrast was significantly higher at 1310 nm than in the visible range.

Two additional advantages were noted in our previous NIR imaging studies over visible light methods. In NIR images of the occlusal surfaces, stains were often not visible, since the organic molecules responsible for pigmentation absorb poorly in the NIR,²⁷ making it easier to identify areas of demineralization. Mild developmental defects²⁸ and shallow demineralization²⁶ appeared differently from deeper, more severe demineralization due to caries, suggesting that we may be able to gauge the severity of lesions by analyzing both NIR reflective images and NIR transillumination images of these surfaces.

Our previous NIR imaging studies utilized InGaAs focal plane arrays (FPAs) that were of low pixel density (25- μm pixel pitch) and an array size of 320×240 pixels. InGaAs FPAs are very expensive and challenging to manufacture. A new NIR imaging technology, the germanium-enhanced complementary metal-oxide semiconductor (CMOS), can potentially be less expensive and more practical for low-cost, high-volume manufacturing. Moreover, the NIR CMOS camera had a larger array size (640×240) and higher pixel density (10- μm pixel pitch).

In summary, the objective of this imaging study was to determine if the area and contrast of occlusal caries lesions in NIR images can be used to estimate lesion severity, and to compare the image contrast of natural caries lesions measured using two NIR imaging technologies.

2 Materials and Methods

2.1 Sample Preparation

Extracted human teeth with natural occlusal caries lesions ($n=99$) were collected from patients in the San Francisco Bay area with approval from the University of California, San Francisco (UCSF) Committee on Human Research, cleaned, sterilized with gamma radiation, and stored in a 0.1% thymol solution to preserve tissue hydration and to prevent bacterial growth. Radiographs and visible light images were acquired of each tooth. Radiographs were acquired directly on Ultra-Speed™ F-speed film (Kodak, Rochester, New York) using 75 kVp, 15 mA, and 12 impulses.

2.2 Near-Infrared Imaging

Two different NIR imaging devices operating at 1310 nm were used to capture transillumination images. The first device used was a high sensitivity InGaAs FPA, model SU320KTSX from Sensors Unlimited (Princeton, New Jersey). The second device used was a Ge-enhanced CMOS image sensor, model NP-EC700M-01 evaluation camera, from NoblePeak Vision (Wakefield, Massachusetts). NIR light was provided by a $\lambda=1310$ -nm superluminescent diode (SLD) with an output power of 15 mW and a 35-nm bandwidth, model SLED1300D20A from Optospeed (Zurich, Switzerland). NIR images were taken using the configurations shown in Fig. 1. Two imaging configurations were used for transillumination. In the first setup, the output from a single-mode optical fiber was collimated by a 20-mm-diam NIR fiber collimator from μLS Micro Laser Systems (Garden Grove, Cali-

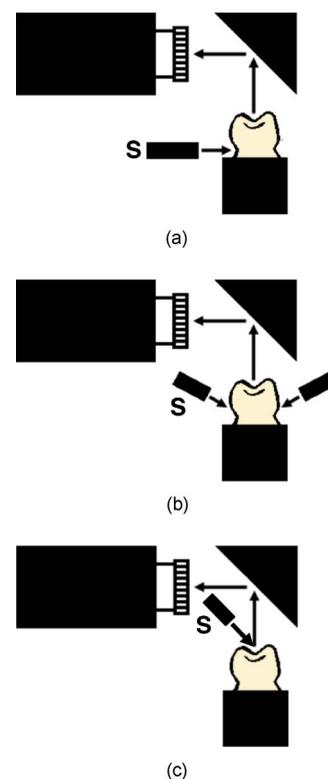


Fig. 1 Different NIR imaging configurations used. (a) NIR 1310-nm source (S) focused with a cylindrical lens on the CEJ. (b) Two sources on each side of the tooth directed at the CEJ. (c) Reflectance imaging in which the NIR light is directed at the tooth occlusal surface. (Color online only.)

fornia) and focused by a 150-mm focal length cylindrical lens at a position just above the cementum-enamel junction (CEJ) of each tooth [Fig. 1(a)]. A sheet of light was delivered to the CEJ. This light serves as a subsurface illumination source that propagates up through the tooth and through the enamel of the occlusal surface, which is imaged by the NIR imaging systems. This is the same setup that we used in previous *in vitro* occlusal imaging studies.²⁷ The second setup utilized the novel arrangement that we previously used for *in vivo* imaging,²⁹ incorporating two fibers with Teflon optical diffusers aimed at a position just above the CEJ of each tooth [Fig. 1(b)]. This second setup was only used with the InGaAs FPA.

NIR reflectance images were acquired using only the InGaAs FPA [Fig. 1(c)]. For reflectance measurements, the surface was illuminated without the cylindrical lens, and crossed polarizers were used to remove the specular reflection from the tooth surface (glare).

2.3 Polarized Light Microscopy

The teeth were mounted on orthodontic resin blocks that are used for both imaging and for histological sectioning. Sections 200 μm thick were cut using a linear precision saw, the IsoMet 5000 (Buehler, Lake Buff, Illinois). Polarized light microscopy (PLM) was carried out using a Meiji Techno RZT microscope (Saitama, Japan) with an integrated digital camera, the Canon EOS Digital Rebel XT from Canon (Tokyo,

Japan). The sample sections were imbibed in water and examined in the brightfield mode with cross-polarizers and a red I plate with 550-nm retardation.

2.4 Transverse Microradiography

Thin sections used in PLM were also imaged using transverse microradiography (TMR). A custom-built digital TMR system was used to measure mineral loss in the different partitions of the sample.³⁰ A high-speed motion control system with a Newport (Irvine, California) UTM150 and 850G stages, and an ESP300 controller coupled to a video microscopy and laser targeting system was used for precise positioning of the tooth samples in the field of view of the imaging system. The volume percent mineral for each sample thin section was determined by comparison with a calibration curve of x-ray intensity versus sample thickness, created using sound enamel sections of 86.3 ± 1.9 vol.% mineral varying from 50 to 300 μm in thickness. The calibration curve was validated via comparison with cross sectional microhardness measurements. The volume percent mineral determined using microradiography for section thickness ranging from 50 to 300 μm correlated highly with the volume percent mineral determined using microhardness $r^2=0.99$.³⁰ TMR was primarily used in this study to determine the penetration of the lesion through the dentin. It is easy to measure the extent of the lesions in enamel using PLM; however, it is much harder to differentiate lesions in dentin using PLM, and TMR was needed to confirm the depth of demineralization.

2.5 Histology and Image Analysis

Lesions were examined and classified according to the lesion depth as follows: (E1) less than halfway to the dentin-enamel junction (DEJ), (E2) more than halfway to the DEJ, (D1) penetration to the DEJ but less than halfway to the pulp chamber, and (D2) more than halfway through the dentin into the inner dentin adjacent to the pulp chamber.

Line profiles with a width of five pixels were extracted from each NIR image aligned with the histological sections, and the lesion or image contrast was calculated using $(I_S - I_L)/I_S$; where I_S is the mean intensity of the sound enamel, i.e., the mean of all the pixels from the sound enamel along the line profile, and I_L is the mean intensity from the corresponding lesion portion of the line profile. The line profiles match the position of the thin sections cut for examination using PLM and TMR. The image contrast varies from 0 to 1, where 1 is very high contrast and 0 is no contrast. Figure 2 shows visible and NIR images of one of the lesions used in the study. The dotted line demarcates the position of the line profile used for calculation of the contrast, and the line matches the corresponding PLM image of the thin section that is also shown. The contrast is reversed for reflectance measurements, i.e., $(I_L - I_S)/I_L$. The number of pixels in the lesion area was also measured using image analysis software, both in reflectance and in transmission. All image analyses were carried out using Igor Pro software from Wavemetrics (Lake Oswego, Oregon). A one-way analysis of variance (ANOVA) followed by the Tukey-Kramer posthoc multiple comparison test was used to compare the lesion contrast and lesion area for all groups for each type of lesion employing InStat soft-

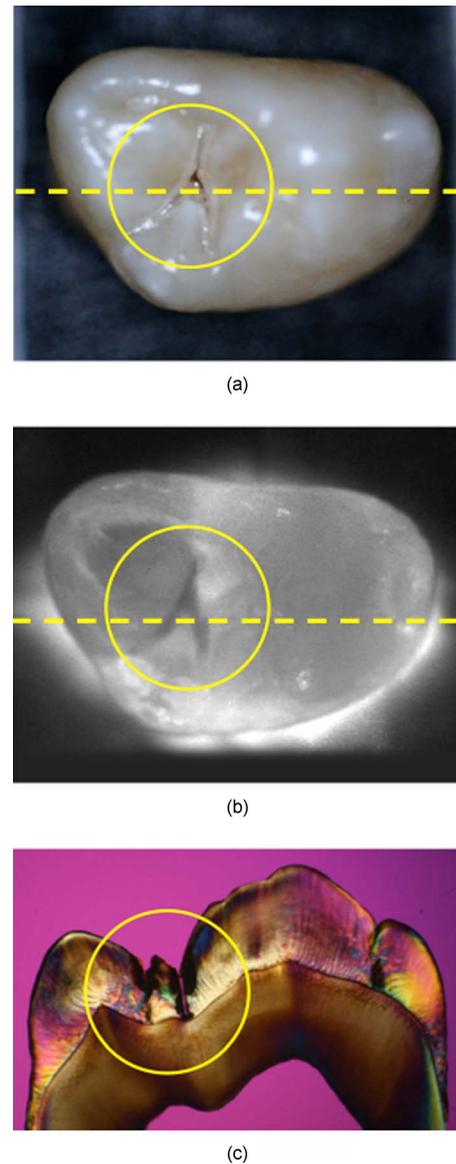


Fig. 2 (a) Visible-light reflectance and (b) NIR transillumination images. The NIR image was captured with the Ge-enhanced CMOS camera using the setup of Fig. 1(a). (c) A PLM image of a thin section cut along the yellow dotted lines in (a) and (b) shows the position and depth of the lesion that penetrates to the dentin-enamel junction and is classified as a D1 lesion. (Color online only.)

ware from GraphPad (San Diego, California). Only one lesion per tooth was used in the analysis.

3 Results

Several images of a third molar tooth with a deep lesion (D2) that were taken using visible light reflectance and NIR transillumination using two NIR imaging setups are shown in Fig. 3, along with a radiograph and PLM images of two sections that were cut along the yellow dotted lines, as indicated in Fig. 3(a). The visible light reflectance image in Fig. 3(a) manifests staining and discoloration; however, there is no decay visible in the radiograph. Therefore, the conventional methods of examination do not reveal the severity of the oc-

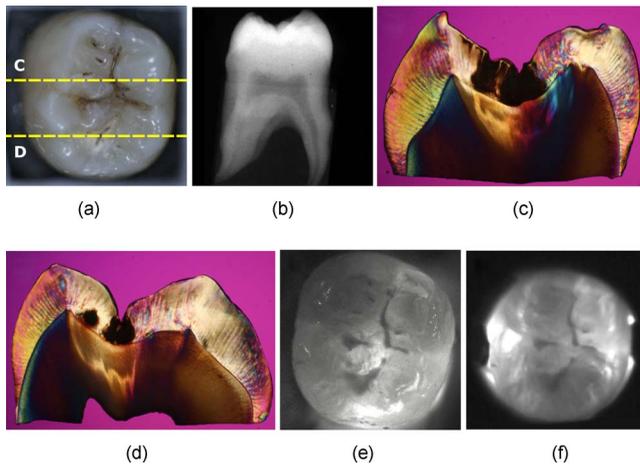


Fig. 3 Images of one of the tooth samples with a D2 lesion. (a) Visible-light reflectance image, (b) radiograph, (c) and (d) PLM images of the sections cut along the yellow dotted lines in (a). NIR transillumination images captured with the (e) Ge-enhanced CMOS camera [setup of Fig. 1(a)] and (f) NIR InGaAs FPA [setup of Fig. 1(b)]. The lesion areas appear as darker areas in the NIR images. These are best resolved in (e). The lesion areas are the dark areas in the enamel in the PLM images (c) and (d). The lesion areas are harder to resolve in the dentin using PLM, and they actually appear lighter than the sound dentin. Therefore, TMR was used to confirm the lesion depth in dentin. (Color online only.)

clusal decay. Typically, occlusal lesions are not visible in a radiograph unless they are very severe, and the D2 lesion in this tooth is not visible on the radiograph, nor are the less severe lesions shown in Figs. 2, 4, and 5. The histological sections of Figs. 3(c) and 3(d) indicate that the lesions penetrate into the dentin. The NIR transillumination images shown in Figs. 3(e) and 3(f) show the lesion areas with high contrast without the interference of staining and discoloration, and these areas of high contrast match the positions of deep lesion penetration indicated in the histology. Staining and dis-

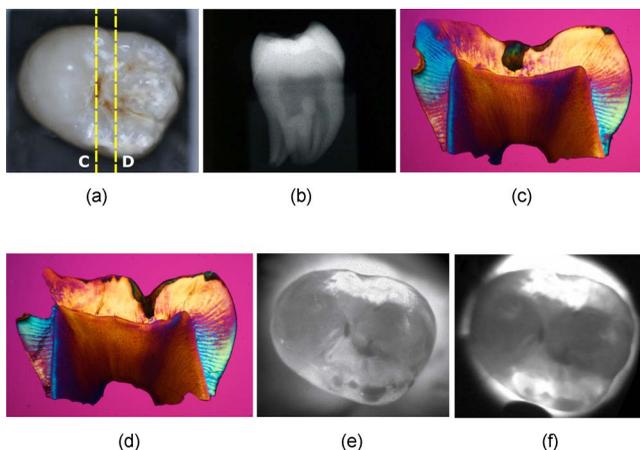


Fig. 4 Images of one of the tooth samples with an E2 lesion. (a) Visible-light reflectance image, (b) radiograph, (c) and (d) PLM images of the sections cut along the yellow dotted lines in (a). NIR transillumination images captured with the (e) Ge-enhanced CMOS camera [setup of Fig. 1(a)] and (f) NIR InGaAs FPA [setup of Fig. 1(b)]. (Color online only.)

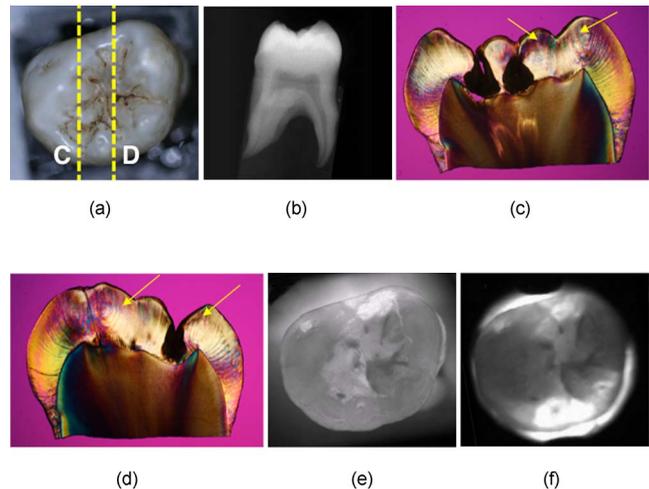


Fig. 5 Images of one of the tooth samples that had a D1 lesion with extensive hypomineralization. (a) Visible-light reflectance image, (b) radiograph, (c) and (d) PLM images of the sections cut along the yellow dotted lines in (a). NIR transillumination images captured with the (e) Ge-enhanced CMOS camera [setup of Fig. 1(a)] and (f) NIR InGaAs FPA [setup of Fig. 1(b)]. The yellow arrows in the PLM images indicate the position of hypomineralization. (Color online only.)

coloration can interfere with imaging in the visible region; however, the chromophores responsible for stains do not absorb NIR light and therefore do not interfere at NIR wavelengths, as can be seen by comparing the NIR images of Figs. 3(e) and 3(f) with Fig. 3(a). Polarized light microscopy images of the two cross sections are also shown in Figs. 3(c) and 3(d). Dark regions in the PLM images of enamel indicate areas of demineralization, and they are easy to distinguish from the transparent sound regions. However, it is more difficult to differentiate dentin lesions from sound dentin using PLM alone. For the samples that were difficult to classify by using PLM, TMR was used to verify the severity of the lesion.

Images from another tooth with less severe lesions, confined to the enamel are shown in Fig. 4. The lesions are clearly visible in the NIR images, and their position matches the PLM micrographs of the two thin sections. Figure 5 shows another tooth with lesions that penetrate to the DEJ (D1). This tooth is interesting, because in addition to the D1 lesions, large areas of the occlusal surface appear with opposite contrast, that is, they appear whiter or with higher intensity than the sound enamel areas. We have observed this phenomenon in previous transillumination images of shallow artificially demineralized areas in the occlusal surfaces,²⁶ and from shallow developmental defects.²⁸ The PLM micrographs show a dark (brown) band of hypomineralization near the enamel surface (see yellow arrows) with an outer layer of higher mineral content that is transparent, which is typical of mild developmental defects or fluorosis.²⁸ We postulate that these areas with defects appear whiter, since the scattering of NIR photons incident at oblique angles that would normally not be emitted from those surfaces, produces an increased NIR photon flux from those surfaces. Namely, many of the NIR photons that are incident on the occlusal surface of the tooth have a high angle of incidence that would typically result in internal reflection; however, the increase in scattering at the defects lo-

Table 1 Table of values for mean contrast ratio and lesion area (transmission) ($n=47$ lesions). Groups with the same depth category listed are statistically similar $p>0.05$.

Depth category	Number of lesions	Mean (SD) contrast reflection	Mean (SD) contrast transmission	(C_T/C_R) mean contrast ratio (SD)	Mean (SD) lesion area transmission	Mean (SD) lesion area reflection
E	19	0.17(0.085) D1, D2*	0.12(0.054) D1	0.75 (0.29) D1	201 (164) D1	213 (225) D1
D1	16	0.17(0.065) E, D2	0.18(0.062) E	1.14 (0.32) E, D2	288 (289) E	294 (208) E
D2	12	0.21(0.10) E, D1	0.36(0.11)	1.98 (1.11) D1	561 (447)	908 (931)

calized to the enamel surface results in an increase in photon flux from those surfaces. The whiter surface also appears more opaque than the sound enamel due to that highly scattering layer. Note the difference in appearance between the outer sound part of the tooth where the sound enamel appears whiter due to directly transmitted light (saturation) and the defect areas that appear more opaque.

The mean contrast values are shown for reflection and transillumination in Table 1. These measurements were taken using the InGaAs FPA in the imaging configurations shown in Figs. 1(a) and 1(c). The lesion contrast in the NIR reflectance images did not increase significantly with lesion severity score (depth) ($p>0.05$), which was determined using PLM and TMR analysis of the histological thin sections. The lesion contrast in transillumination increased with lesion depth. The contrast of the severe D2 lesions was significantly higher than the less severe lesion groups ($p<0.01$). The mean contrast of the less severe dentinal lesions (D1) was higher than for the lesions confined to enamel (E); however, they did not break statistically ($p>0.05$). The ratio of contrast in transillumination over reflection was also calculated to determine if this value provided greater separation between the lesions. However, this parameter did not provide a significant improvement over transillumination alone. There was still a significant difference between the deep dentinal lesions (D2) and the shallower lesions ($p<0.01$), but not between enamel (E) and shallow dentinal lesions (D1) ($p>0.05$), as can be seen in Table 1.

The area of the lesion in both the reflectance and transillumination NIR images was also measured, and the lesion area increased with increasing lesion severity. In reflectance and transillumination, the area measurements of enamel lesions could be differentiated from those of dentinal lesions ($p<0.01$).

Table 2 lists the lesion contrast values measured using the Ge-enhanced CMOS camera and the imaging setup of Fig. 1(a), along with those measured using the InGaAs FPA with the imaging setup of Fig. 1(b). The image contrast for the enamel-only lesions E1 and E2 were both significantly lower ($p<0.05$) than the lesions that penetrated to dentin using the Ge-CMOS setup, although the enamel groups did not break statistically with each other. For the InGaAs FPA, E1 broke with D1 and D2, but E2 did not break statistically with either E1 or D1. However, if the enamel-only lesions are combined into one group (E1+E2), then group (E) does break statistically with both the D1 and D2 groups.

A comparison of the lesion contrast measured for the two different groups of teeth using the InGaAs and Ge-CMOS imaging systems employing the same imaging setup of Fig. 1(a) for transmission indicates that the contrast was significantly higher for E, D1, and D2 lesions ($p<0.001$) for the Ge-CMOS imaging system. A comparison between the two imaging setups for transmission, Figs. 1(a) and 1(b), indicates that the clinical imaging setup, Fig. 1(b), that employed two probes yielded significantly higher contrast for E, D1, and D2 lesions as well $p<0.01$.

4 Discussion

This study indicates that the area and contrast of occlusal lesions measured using NIR transillumination can potentially be used as an indicator of lesion severity. The decision on whether surgical intervention is necessary or more conservative chemical intervention will suffice for the treatment of occlusal lesions is a primary concern of the clinician. Penetration of the lesion deep into dentin is one key indicator for surgical intervention, and the D2 lesions are the most likely lesions to require surgical intervention, since these are the lesions least likely to be arrested by chemical intervention or fluoride treatment. The lesion contrast of the D2 lesions was significantly higher for both imaging systems than the other less severe lesions. Since lesion severity is assigned on the basis of depth, it is important to point out that in many cases there is little difference in the severity of lesions in adjoining

Table 2 Mean contrast ratio for Ge-CMOS and InGaAs FPA imaging systems ($n=52$ lesions). Groups with the same depth category listed are statistically similar $p>0.05$. E only compared with D1 and D2. Groups sharing alphabet letters are statistically similar $p>0.05$.

Lesion category	Number of lesions	Lesion contrast Ge (CMOS)	Lesion contrast (InGaAs FPA)
E1	7	0.30±0.065 E2*	0.26±0.090 E2
E2	15	0.35±0.086 E1	0.34±0.091 E1, D1
E (E1+E2)*	22	0.33±0.081	0.31±0.095
D1	13	0.45±0.059	0.39±0.071 E2
D2	17	0.53±0.086	0.47±0.068

groups. For example, a lesion that is close to the DEJ would be considered an E2 lesion, while if it touches the DEJ it is considered a D1 lesion, even if they are very similar in depth and severity. The highest contrast was achieved for the higher resolution Ge-CMOS camera with 10- μm pixel pitch. It appears that the higher pixel density and larger array size of the Ge-enhanced CMOS camera provided increased contrast of the occlusal lesions. This increase in contrast was sufficient to demonstrate a significant difference in the mean contrast between the lesions confined to enamel (E) and the deeper D1 and D2 dentin lesions. Moreover, the contrast was also significantly higher for the more severe D2 lesions than the D1 lesions. The lesion contrast for the E, D1, and D2 groups also broke statistically for the lower resolution InGaAs FPA when the two-source illumination probe [Fig. 1(b)] was used. The two-source illumination probe illuminates both facial and lingual surfaces of the tooth, producing more uniform illumination across the occlusal surface. This probe worked well in our first *in vivo* clinical study in which we were able to image approximal lesions from the occlusal surface using the InGaAs FPA.²⁹ The ability to achieve uniform illumination across the occlusal surface is key for the discrimination of lesions and to avoid false positives. Another method of avoiding false positives created by the illumination geometry is to acquire images from multiple views by changing the angle of the illumination probes. In future studies we will investigate the performance of the two source probe of Fig. 1(b) with the Ge-CMOS camera. The Ge-CMOS camera was a prototype that was too large to be feasible for clinical use, while the InGaAs FPA system has been used for *in vivo* imaging.

Although the NIR reflectance measurements did not exhibit higher contrast with increasing lesion depth in this study, a previous study demonstrated that NIR reflectance measurements are useful for the detection of early demineralization.²⁶ Polarized NIR provided improved contrast over polarized visible reflectance measurements for areas of shallow demineralization on enamel surfaces.

Since the contrast for transillumination is sometimes reversed for shallow demineralization or minor developmental defects (fluorosis), as demonstrated in Fig. 5, we anticipated that the combination of NIR images in transmission and reflection would provide better discrimination of lesion depth. However, that was not the case for the severe lesions investigated in this study. This approach is most likely to be successful for very shallow lesions, and we plan to further investigate this approach using different models of early demineralization.

The ability to discriminate between staining and fluorosis and deeply penetrating caries lesions, as demonstrated by Fig. 5 and in prior NIR imaging studies, is an important advantage.^{27,28} Most occlusal lesions are initially detected by visual staining of the fissure, and it is difficult to discriminate staining from demineralization. Previous NIR studies have demonstrated that stains are less visible in the NIR, since the organic molecules responsible for pigmentation absorb poorly in the NIR, making it easier to identify areas of demineralization.²⁷ This should be particularly advantageous for *in vivo* screening for caries, since the interference from staining and hypomineralization is likely to be greatly reduced.

This *in vitro* study demonstrates that increasing lesion con-

trast and lesion area in NIR images corresponds with increasing depth and severity of occlusal caries lesions. The next step is to acquire *in vivo* NIR imaging of occlusal caries lesions, which is planned for the near future.

Acknowledgments

The authors acknowledge the support of NIH grant R01-DE14698. The authors would like to thank Noble Peak vision for loan of the Ge-enhanced CMOS camera.

References

1. J. Barenie, G. Leske, and L. W. Ripa, "The use of fiber optic transillumination for the detection of proximal caries," *Oral Surg.* **36**, 891–897 (1973).
2. C. M. Pine, "Fiber-optic transillumination (FOTI) in caries diagnosis," *Early Detection Dental Caries*, pp. 51–66 (1996).
3. J. Peltola and J. Wolf, "Fiber optics transillumination in caries diagnosis," *Proc. Finn Dent. Soc.* **77**, 240–244 (1981).
4. R. D. Holt and M. R. Azevedo, "Fiber Optic transillumination and radiographs in diagnosis of approximal caries in primary teeth," *Community Dent. Health* **6**, 239–247 (1989).
5. C. M. Mitropoulos, "The use of fiber optic transillumination in the diagnosis of posterior approximal caries in clinical trials," *Caries Res.* **19**, 379–384 (1985).
6. H. Hintze, A. Wenzel, B. Danielsen, and B. Nyvad, "Reliability of visual examination, fibre-optic transillumination, and bite-wing radiography, and reproducibility of direct visual examination following tooth separation for the identification of cavitated carious lesions in contacting approximal surfaces," *Caries Res.* **32**(3), 204–209 (1998).
7. A. Schneiderman, M. Elbaum, T. Schultz, S. Keem, M. Greenebaum, and J. Driller, "Assessment of dental caries with digital imaging fiber-optic transillumination (DIFOTI): in vitro study," *Caries Res.* **31**, 103–110 (1997).
8. M. Bin-Shuwaish, P. Yaman, J. Dennison, and G. Neiva, "The correlation of DIFOTI to clinical and radiographic images in Class II carious lesions," *J. Am. Dent. Assoc.* **139**(10), 1374–1381 (2008).
9. D. A. Young and J. D. Featherstone, "Digital imaging fiber-optic transillumination, F-speed radiographic film and depth of approximal lesions," *J. Am. Dent. Assoc.* **136**(12), 1682–1687 (2005).
10. D. A. Young, "New caries detection technologies and modern caries management: merging the strategies," *Gen. Dent.* **50**(4), 320–331 (2002).
11. J. Yang and V. Dutra, "Utility of radiology, laser fluorescence, and transillumination," *Dent. Clin. North Am.* **49**(4), 739–752 (2005).
12. S. Keem and M. Elbaum, "Wavelet representations for monitoring changes in teeth imaged with digital imaging fiber-optic transillumination," *IEEE Trans. Med. Imaging* **16**(5), 653–663 (1997).
13. D. Fried, J. D. B. Featherstone, R. E. Glana, and W. Seka, "The nature of light scattering in dental enamel and dentin at visible and near-IR wavelengths," *Appl. Opt.* **34**(7), 1278–1285 (1995).
14. R. S. Jones and D. Fried, "Attenuation of 1310-nm and 1550-nm laser light through sound dental enamel," in *Lasers in Dentistry VIII*, *Proc. SPIE* **4610**, 187–190 (2002).
15. C. L. Darling, G. D. Huynh, and D. Fried, "Light scattering properties of natural and artificially demineralized dental enamel at 1310-nm," *J. Biomed. Opt.* **11**(3), 034023: 1–11 (2006).
16. G. D. Huynh, C. L. Darling, and D. Fried, "Changes in the optical properties of dental enamel at 1310-nm after demineralization," in *Lasers in Dentistry X*, *Proc. SPIE* **5313**, 118–124 (2004).
17. C. L. Darling and D. Fried, "Optical properties of natural caries lesions in dental enamel at 1310-nm," in *Lasers in Dentistry XI*, *Proc. SPIE* **5687**, 102–110 (2005).
18. G. Jones, R. S. Jones, and D. Fried, "Transillumination of interproximal caries lesions with 830-nm light," in *Lasers in Dentistry X*, *Proc. SPIE* **5313**, 17–22 (2004).
19. R. S. Jones, G. D. Huynh, G. C. Jones, and D. Fried, "Near-IR transillumination at 1310-nm for the imaging of early dental caries," *Opt. Express* **11**(18), 2259–2265 (2003).
20. B. Angmar-Mansson and J. J. ten Bosch, "Optical methods for the detection and quantification of caries," *Adv. Dent. Res.* **1**(1), 14–20 (1987).

21. J. J. ten Bosch, H. C. van der Mei, and P. C. F. Borsboom, "Optical monitor of in vitro caries," *Caries Res.* **18**, 540–547 (1984).
22. P. E. Benson, A. Ali Shah, and D. Robert Willmot, "Polarized versus nonpolarized digital images for the measurement of demineralization surrounding orthodontic brackets," *Angle Orthod.* **78**(2), 288–293 (2008).
23. M. J. Everett, B. W. Colston, U. S. Sathyam, L. B. D. Silva, D. Fried, and J. D. B. Featherstone, "Non-invasive diagnosis of early caries with polarization sensitive optical coherence tomography (PS-OCT)," in *Lasers in Dentistry V*, *Proc. SPIE* **3593**, 177–183 (1999).
24. D. Fried, J. Xie, S. Shafi, J. D. B. Featherstone, T. Breunig, and C. Q. Lee, "Early detection of dental caries and lesion progression with polarization sensitive optical coherence tomography," *J. Biomed. Opt.* **7**(4), 618–627 (2002).
25. D. Fried, J. D. B. Featherstone, C. L. Darling, R. S. Jones, P. Ngaotheppitak, and C. M. Buehler, *Early Caries Imaging and Monitoring with Near-IR Light*, W. B Saunders Co., Philadelphia (2005).
26. J. I. Wu and D. Fried, "High contrast near-infrared polarized reflectance images of demineralization on tooth buccal and occlusal surfaces at $\lambda=1310\text{-nm}$," *Lasers Surg. Med.* **41**, 208–213 (2009).
27. C. M. Bühler, P. Ngaotheppitak, and D. Fried, "Imaging of occlusal dental caries (decay) with near-IR light at 1310-nm," *Opt. Express* **13**(2), 573–582 (2005).
28. K. Hirasuna, D. Fried, and C. L. Darling, "Near-IR imaging of developmental defects in dental enamel," *J. Biomed. Opt.* **13**(4), 044011:1-7 (2008).
29. M. Staninec, C. Lee, C. Darling, and D. Fried, "In vivo near-IR imaging of approximal dental decay at 1310 nm," *Lasers Surg. Med.* **42**(4), 292–298 (2010).
30. C. L. Darling, J. D. B. Featherstone, C. Q. Le, and D. Fried, "An automated digital microradiography system for assessing tooth demineralization," in *Lasers in Dentistry XV*, *Proc. SPIE* **7162**, 71620T (2009).